Review

Effect of Aging on the Immune System

Yoshimura Fukazawa and Keiko Kagaya

Department of Microbiology, Yamanashi Medical College, Tamaho-cho, Yamanashi 409-38, Japan

Abstract: Host defense mechanisms against microbial and viral infections as well as neoplastic cells constitute a close network, with phagocytes, natural killer (NK) cells, immunocytes including T and B cells, and cytokines released from and interacting with these cells. Although aging is accompanied by many changes in the immune system, it is unlikely that all immune cells and systems age at equal rates. Suffice to say, involution of the thymus plays one of major roles in immune senescence. Related to this event are the altered number and functions of T cell subpopulations involved in immunoregulation, a decrease in the immune response by both cell-mediated and humoral branches of the immune system, and an increase in autoimmune activity. The clinical implications of these changes are the elderly person's increased susceptibility to infections such as pneumococcal pneumonia, influenza and tuberculosis. Other changes include an increased susceptibility to neoplasms and perhaps acceleration of the aging process by increased autoimmune activity and immune complexes. The functions of macrophages, PMN, NK cells, and also the complement system are not seriously impaired with age. The evidence that lymphoid progenitors in bone marrow cells from young animals are able to differentiate into lymphoid cells in the aging animal, and that involution of thymus may be restored by manipulation of the endocrine system, suggest that there may be a potential for reconstitution of some immune defects in aged individuals by grafting or treatment with drugs to control various age-related diseases, including cancer.

Key words: Aging, B cells, T cells, Thymus

Introduction

Aging is accompanied by characteristic structural and functional alterations in many organs and systems, with the alterations in the immune system being the most pronounced.

Age-associated changes in the immune system have been studied in humans and in experimental animals, and the effect of increase in age on the immune system has been clearly documented (1–6).

Immune senescence in the elderly is usually associated with an increased susceptibility to infections, neoplasia, autoimmune disorders and vascular disease. This hypothesis has led to the investigation of the methodology by which

Received, December 11, 1989

Accepted, December 14, 1989

immune system dysfunction might be prevented or restored in an effort to delay the inevitable consequence of aging and agerelated disease. The aim of this article is to give an outline of very recent pertinent conclusions concerning the age-related changes in the wide range of the immune system related to the host defense network, as drawn from both human and animal studies. As an introduction, a brief overview of the immune network for host defense will be mentioned.

THE IMMUNE NETWORK FOR HOST DEFENSE

In the past decade immunologic research has revealed the incredibly complex nature of the interaction between cells involved in host defense and invading microorganisms. It is simplistically devided into two categories. The first defense is responsible for the control of infections caused by intracellular pathogens and tumors, and rejection on grafts, in terms of cell-mediated immunity. The second is involved in elimination of extracellular organisms in terms of humoral immunity (7).

Killing of prototypical intracellular bacteria such as Listeria monocytogenes and the mycobacteria, a number of fungal organisms, certain groups of viruses, and parasites is dependent upon the integrated activity of T lymphocytes and macrophages. Recent studies have defined the roles of various cytokines - interleukin 1 (IL-1) (8), interleukin 2 (IL-2) (9), and γ interferon (IFN- γ) (10, 11) - in the process leading to macrophage activation or cellmediated immunity. It has also become increasingly clear that cytotoxic T lymphocytes (CTL) (12, 13) and a separate group of lymphocytes, natural killer (NK) cells (14, 15), participate in defense against tumor cells, certain viruses, and virus-infected cells.

Immunoglobulins and complement factors are the principal elements of humoral immunity, and while a major function of these proteins is the opsonization or killing of extracellular bacteria, they also contribute importantly to defense against viruses and the injurious effects of microbial toxins. Although polymorphonuclear leukocytes (PMN) play a key role in the killing of opsonized bacteria, in the primary (normal) stage of defense, monocytes and macrophages can also eliminate these organisms in the activated stage (11, 16, 17, 18).

To understand the effects of aging on the immune system and its function, the interactions between cells involved in host defense and invading microorganisms and other cells are diagramatically summarized in Figure 1.

AGE-RELATED CHANGES IN THE IMMUNE SYSTEM

Thymus

The work of Good, Miller, and Waksman in the early 1960s revealed the crucial role of the thymus in the immune system. Numerous clinical, biological, and biochemical observations underline the close relations between aging and immunity. In particular, the T cell system and its functions are subjected to age-dependent decline and impairment. The senescence of peripheral immune function is paralleled by the involution of the central organ of the T cell system, the thymus (19).

The involution of the thymus with age is characterized by a puberty-independent continuous degeneration of the thymic epithelial space (20). It starts in the very first years of life and exhibits a constant velocity during the first decade. The velocity of involution decreases progressively. Remnants of thymic epithelial tissue with a cortical lymphocyte population are preserved beyond 100 years of age (21). Thymic atrophy in the aged involves various types of disorganization of individual lobules with T and B lymphocytes often located outside rather within epithelial remnants. The cause of involution and its impact on the immune status of the aged are far from being understood and remains the subject of speculation. The idea that the age-related involution of the immune apparatus and, in particular, the thymus, may be an adaptive mechanism to protect against autoimmune reactions has already been expressed (22).

Lethally irradiated and thymectomized young animals have been reconstituted with bone marrow cells and thymus grafts from donors of varying ages (23, 24). Thymus grafts from newborn animals permit the most rapid reconstitution of the T lymphocyte population and the most complete recovery of responsiveness to T cell mitogens and to T-cell dependent antigens. When thymus grafts are taken from older animals, the pace of recovery is delayed, and in many cases the level of thymusdependent immune function never reaches that seen in intact animals or in animals reconstituted with neonatal thymus glands. Thus, the capacity of the thymus to affect the maturation of immature T lymphocytes de-

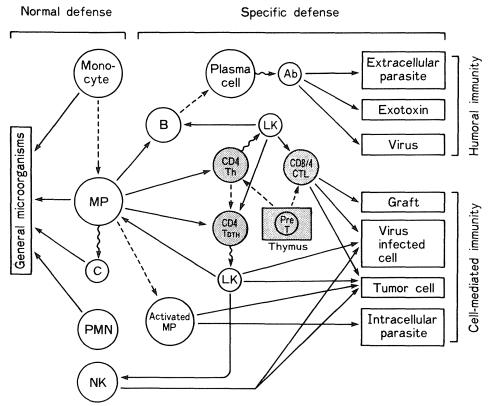


Fig. 1. Schematic diagram of the cellular and molecular interactions in host defense network.

—, interaction; ---, differentiation; ~--, secretion. Ab, antibody; LK, lymphokine; C, complement; shaded parts, sites that are most vulnerable to aging.

creases with age.

The level of thymic hormones in the serum of humans and experimental animals begins to fall soon after the morphologic involution of the thymus gland. Thus, in humans between the ages of 20 and 30, the serum level of thymic hormone begins to fall, and after the age of 60, thymic hormone is no longer detectable in serum (25).

It has been found that thymosin can overcome deficient T helper cell activity and improve *in-vitro* antibody responses of human lymphocytes to influenza vaccine (26), probably as a result of a thymosin-mediated increase in production of IL-2 (27). In another study, thymostimulin, a bovine thymic extract, has been administered parenterally for 3 months to aged hospitalized patients, and

although no obvious changes were seen in measured immunologic parameters, patients in the treated group had significantly fewer infections than controls (28).

The thymus, which is grossly atrophied in 12 to 15-month-old male rats, is markedly restored in size 30 days after orchidectomy. The organ then appears normal histologically, having a well-defined cortex and medulla, is vascularized and filled with thymocytes. The regeneration of the thymus after orchidectomy was inhibited in a dose-related fasion by testosterone implants which produced serum concentrations of testosterone within the physiological range. The thymus also increased in size after orchidectomy of 10-week-old rats, and testosterone inhibited the enlargement of the thymus (29). The effects of several steroids

on the regenerating thymus in aging male rats have been studied (30). The results showed the possibility that testosterone and oestradiol may have caused atrophy of the thymus, while 5α -dihydroxy-testosterone may have retarded regeneration of the thymus without any atrophic effects.

Although it is proposed that involution of the thymus gland during the first half of the life span is followed by marked alterations in immune function, cell-mediated immunity and T-cell dependent humoral immune response still remain, if partially, in the aged. The mechanism by which decreased remnant mature lymphocytes in the aged still hold immune function is not clear.

T cells

Contradictory results have been reported regarding the number and proportion of T cells and their subpopulations in the elderly. Regarding the absolute number of peripheral blood, lymphocytes show relatively by constant (31, 32) or little change (33), a decrease in the circulating T lymphocytes (34, 35, 36), and an increase during aging (37). With regard to subpopulations of T lymphocytes, the number of immature T cells is increased in the elderly (38, 39). The number of T helper cells OKT4⁺ (CD4⁺) has been reported to be increased (40, 41) or unchanged (41, 42) with age, while the number of T suppressor cells OKT8⁺ (CD8⁺) has been reported to be increased (32, 41), decreased (38, 42) or constant (39, 43). Both proportions and absolute numbers of OKT3+ (CD3, pan T cell marker) and OKT4+ (CD4) cells were reduced in the elderly as opposed to those in young control population (44).

By studying 206 apparently healthy aged individuals, a slight decrease in the frequency of T4⁺ (CD4) and T8⁺ (CD8) cells has been found (45). Furthermore, these data are also supported by the demonstration that the number of null cells (non-T, non-B lymphocytes) is increased in the peripheral blood of aged individuals (46). Much of the controversy may

be accounted for by differences in the technology employed or failure to define properly the subject populations. Regarding the effect of gender, elderly women displayed T3⁺ (CD3), T4⁺ and T8⁺ cell numbers comparable to those seen in young women. Elderly men exhibited a reduction of T3⁺ and T4⁺ lymphocytes when compared with either young men or women (44).

Although there is no reduction in the ability of T lymphocytes from the elderly to bind lectin such as phytohaemagglutinin (PHA), PHA-inducible lymphocyte activation declines with age (47, 48). This seems to be due in part to a reduction in the number of PHA responsive lymphocytes and in part to a reduction in the number of sequential cell divisions occuring in lympocytes from elderly vs. young subjects (49). The poor mitogen response of mononuclear cells from elderly volunteers correlated with an increased sensitivity to prostaglandin E₂ (50); this increase was reversed in vitro by the addition of the arachidonic cyclo-oxygenase inhibitor, indomethacin, and by lithium carbonate (50, 51). Additional studies in aged experimental animals have also revealed a decrease in the response of T cells to nonspecific mitogens (39, 52, 53).

Antigen-specific or antigen-nonspecific suppressor cells have received considerable attention in immunogerontology because of their important role in regulating the immune response as well as the induction and maintenance of tolerance to exogenous and self antigens (54, 55). Suppressor cell activity has been found increased (56–58), unchanged (56) or decreased (59-61) in aging mice and humans. The overt controversy as to age-related changes in immunosuppression reflects the large variety of methods used to assess suppressor cell activity. More recently, Doria et al (62) reported age-related alterations of antigen-specific T cell-mediated suppression in the 4-hydroxy-3-nitrophenyl acetyl (NP) system, suggesting that aging may affect the recognition repertoire expressed in suppressor T cell

subsets (inducer, transducer and effector suppressor T cells). Moreover, the finding that suppression is less efficient when exerted up on spleen cells from old rather than from young mice provides an explanation for the increased frequency of autoimmune disorders in aging. Studies on in vitro induction of Con A-activated T helper cells, T suppressorinducer cells and T suppressor cells from aged Peyer's patches (PP) indicated that the generation of T suppressor cells was largely impaired, in contrast to minor defect(s) in that of T helper cells (63), suggesting that in aged PP, a T suppressor-inducer cell subset appears to be more selectively impaired during the aging process than the other lymphocyte subpopulations. On the other hand, the ability to generate suppression to newly encountered antigen declines with age, whereas a resident splenic suppressor cell population accumulates over the lifetime of the animals (64).

One hypothesis suggests that defects in the capacity of T cells to produce or respond to T cell growth factor, or IL-2, may be the fundamental cause of the immune deficiency seen with aging. Most evidence from studies of humans supports a decrease in IL-2 production with aging (40, 65, 66). Data from animal studies also support a decrease in IL-2 production with aging (67-69), although one study revealed no difference between IL-2 production in elderly rats and that in young rats (39). Investigations regarding the response of T cells from elderly humans to exogenous IL-2 have given contradictory results, showing intact response (70), or defective responses (40, 65). Similarly, conflicting results have been obtained in studies of the T cell response from aged animals to exogenous IL-2 (39, 60, 67). Further investigations are needed to define the exact role of interleukins in the pathogenesis of immune defects associated with aging.

The level of IFN- γ , another lymphokine thought to be of importance in immunoregulation, resistance to viral infections and also in macrophage activation (11, 16–18), was re-

portedly normal (71) or decreased (72) in response to antigen or mitogen in elderly subjects. Other investigators reported an age-associated decline in the synthesis and secretion of both IFN- γ and IFN- α by mononuclear cells (73). Whether dificiencies in these lymphokines result in the increased susceptibility of aged humans to viral infection and malignant disease remains to be determined.

Investigation regarding the genetic analysis of Con A-stimulated cultures of spleen cells from old and young mice has been reported to show that aging led to a consistent decline in the level of *c-myc* mRNA in stimulated cells suggesting that these deficits may involve, at least for some gene, alterations in post-transcriptional processing (74).

B cells

Unlike conflicting data on T cell proportion during aging, relative frequency of B lymphocytes is generally unaffected in the elderly, in spite of increased or decreased functional capacities (75). By contrast, an investigation revealed that aged mice are impaired in their ability to generate B cells and that this may be caused in part by a reduction in the frequency of pre-B cells, as well as to a lack of support for these cells in the process of their maturation to B cells (76).

Polyclonal immunoglobulin (Ig) response to pokeweed mitogen (PWM) in a group of aged individuals was either normal or increased (32). By contrast, antigen-specific and polyclonal responses with advancing age was reported to be reduced (77). Antigen-specific response was more reduced than polyclonal response (77). Furthermore, an intrinsic defect of B cell maturation in the elderly has been also postulated (78), which has been confirmed by recent findings showing that both IL-1 and IL-2 are able to enhance signficantly the diminished B cell response of elderly subjects (79). In addition, the decreased expression of surface markers (sIgM and sIgD) and the changes in intracellular structure of aged B

cells analyzed by flow microcytofluorometry strongly support an alteration in B lymphocytes even if their relative frequency remains constant (79). These data suggest that immunosenescence of B lymphocytes leads to a perturbation of their functional and/or phenotypic characteristics.

On the other hand, aged purified B lymphocyte preparations respond to antigen as plaque-forming cells (PFCs) similar to those observed in young healthy donors (80). This suggests that a non-B population is involved in the impaired antibody synthesis; i.e. the negative modulation of B cell responses mediated by T cells during the elderly.

In conclusion, several factors impair B lymphocyte response during aging, acting either via T cells or directly on B cells even if the regulatory T cell network is taken into consideration.

Macrophages

Macrophages have intrinsic roles for the presentation of antigens for immune response, and phagocytosis and killing of microorganisms. Most animal studies have indicated that the overall number and function of macrophages are unchanged with age. Specifically, chemotaxis, phagocytosis, and intracellular killing have been found to be normal (51, 81, 82). In one study, although monocytes from aged donors showed a normal chemotactic responsiveness to zymosan-activated serum, the chemotactic activity induced by leukocytederived chemotactic factor and phagocytosis were depressed (83).

While macrophages from older animals were not impaired in their ability to inhibit either the intracellular growth of *Toxoplasma gondii* or the DNA synthesis of tumor cells, the induction of these capacities was delayed (51, 84). One study of macrophages from aged animals revealed diminished phagocytic activity (77, 85); another detected decreased nonspecific tumor cell cytotoxicity (86). An investigation regarding the respiratory burst and

bactericidal activity of alveolar macrophages from adult and senescent mice indicate that the enhanced susceptibility of the senescent host to lower respiratory tract infection cannot be attributed to age-related changes in the nonspecific antimicrobial activity of resident alveolar macrophages (87). The helper function of macrophages from older animals was normal in terms of both T and B cell mitogenesis, the production of plaque-forming cells, and the generation of IL-1 (39). On the other hand, macrophages involved in the production of T cell growth factor, IL-2, from lymphocytes are altered with age (88). The capacity of macrophages of old mice to synthesize IL-1 is also markedly reduced (89, 90).

Polymorphonuclear leukocytes

Polymorphonuclear leukocytes (PMN) represent an important defensive mechanism against infectious agents. Aging does not appear to be associated with granulocytopenia (91). In addition, the adherence of PMN from elderly individuals to nylon fibers is reportedly normal (83) or increased (92, 93). Conflicting data, however, have been reported regarding other PMN functions. Several investigators have demonstrated intact chemotaxis, phagocytosis, and bactericidal activity in the aged (94, 95), whereas others have shown significantly impaired PMN chemotaxis (83, 92, 93). Phagocytosis and the bactericidal activity of PMN were also significantly depressed in several studies of the function of PMN from elderly individuals (83, 92, 93, 96, 97). Specifically, nitroblue tetrazolium reduction (31, 83, 92) and the generation of superoxide anions by PMN stimulated with latex particles (83, 98) were impaired with aging. In addition, PMN chemiluminescence was significantly depressed in one study of individuals exceeding 80 years of age (99). Other investigators, however, reported a specific defect in receptor activation of a key phosphorylating activity (100). Despite these reported defects of PMN from the elderly, there was no association with an increased incidence of bacterial infections (92).

Natural killer cells

Functional alterations in cytotoxic lymphoid cells also occur with aging. These cells are instrumental in defense against virus-infected cells and tumor cells. There is general agreement that the ability to generate natural killer cells (NK) diminishes with age in animal models of senescence (101, 102). However, results in humans have been conflicting, revealing a marginal increase in NK activity in aged males, but not in females (103), no significant change (104, 105), a moderate increase in NK activity (106), and an increase or lack of increase depending on the parameter of expression (107). Other investigations revealed an increase in NK activity in the majority of healthy elderly (>80 years) and a decrease in mitogenic response to PWM (108-110).

Complement system

The complement system comprises the other major group of serum proteins involved in opsonization, and complement levels and functional activity appear to be intact in elderly subjects (94).

The effects of aging on effector cells and molecules involved in normal defense mechanisms are shown in Table 1.

AGE-RELATED CHANGES IN THE IMMUNITY

Cell-mediated immunity

Cell-mediated immunity (CMI) plays an important role in defense against certain infectious agents, in surveillance against cancer, and in immune regulation (111). In aged individuals the susceptibility to infections is increased (112). There is ample evidence that the deterioration of the immune system is related to a decreased function of T cells. This decline in immune function appears to be mainly due to impairment of T helper cell activity (113–116). Helper T cells play an important

Table 1. Effects of aging on effector cells and molecules involved in normal defense mechanisms

Effect	Reference(s)
PMN	
Numbers	→ 91
Functions	↓ 31, 83, 92, 93,
	96-100
	\rightarrow 83, 92–95
Mononuclear phagocytes	
Functions	↓ 77, 83–86, 88
	\rightarrow 51, 81, 82, 87
IL-1 production	↓ 89, 90
The second secon	→ 39
Natural killer cells	1
Number or function	↓ 101, 102
	→ 46, 103–110
Complement levels	→ 94
and function	

 $[\]uparrow$, increased; \rightarrow , unchanged; \downarrow , decreased.

role in the generation of inducer T cells which participate in the delayed-type hypersensitivity (DTH) reactions and inductions of B-cell responses as well as cytotoxic T-cell response. The inductive helper cells can be characterized on the basis of their surface markers. They have Thy-1 surface markers but lack Lyt-2 (CD8) membrane antigens (117).

The changes in the ability to induce B-cell responses and, to a lesser extent, cytotoxic T-cell responses in aging individuals have been described (118-120). The DTH responses represent the capacity of the immune system to cope with various types of infections of intracellular microorganisms, such as Mycobacterium tuberculosis (121), Salmonella typhimurium (122), Listeria monocytogenes (123), and Candida albicans (124). It has been shown that the DTH response to a panel of antigens decreased with increasing age (125). A study employing an experimental mouse model of Mycobacterium tuberculosis infection showed that old mice are more susceptible to M. tuberculosis in that they are unable to survive an infectious dose that is sublethal to young abult mice. Passive transfer of adoptive immunity from mice of increasing age revealed that the increased susceptibility of aged mice is associated with a deficient capacity to generate protective T lymphocytes to the *M. tuberculosis* infection (126).

In the animal model of genetic background, it is well known that C57BL/Ka mice are more sensitive to age-related immune disorders than CBA/Rij mice. C57BL/Ka mice show a relatively high frequency of pathological lesions of the immune system with age (127). In both experimental animals and humans, three stages of susceptibility to viral infections are apparent; the neonatal state, characterized by enhanced susceptibility to infections; childhood and adolescence, during which there is decreased susceptibility; and adulthood (sexual maturity), characterized by increased susceptibility to primary viral infections with advancing age (128). Moreover, advanced age is associated with reactions of latent viruses, most notably varicella zoster virus (VZV) (129), and most likely, oncogenic viruses as well. The mechanisms responsible for these alterations in susceptibility to viral infections have not been completely elucidated. Differences in antibody production do not seem to play a role. Most authors feel that depression of CMI, as measured by delayed cutaneous hypersensitivity or lymphocyte stimulation by mitogens and antigens, may be of importance. Recent studies revealed that in vitro lymphocyte proliferative resopnses to VZV by lymphocytes of adults aged 12-46 years are mainly by CD4+ T cells and that this subset can lyse VZV-infected cells with HLA-DR surface antigens directly (130). An investigation suggested that mononuclear cells capable of killing VZV-infected target cells persist with aging but that reduced numbers of antigen-responsive and lymphokine-releasing T cells may limit their function (131).

An investigation regarding interferon formation in response to coxsackie virus B3 infection in mice suggested that adult mice

produce relatively less interferon in relation to the amount of virus replicated in their tissues than do younger animals (128).

Delayed-type hypersensitivity and graft rejection are two classic manifestations of CMI in vivo. In a system producing acquired immunologic tolerance to an allograft by injecting cells from the donor, survival of the allograft is dependent on several factors that include the dose of tolerogenic cells, antigenic disparity between the recipient and donor, and the developmental stage of the recipient (132). The age of adult recipient mice was found to be crucial to the induction of skin allograft tolerance with allogeneic spleen cells plus cyclophosphamide. By contrast, the age of the donor mice used for tolerance induction did not appear to be crucial for the induction of a tolerant state (133).

The ability to mount a mixed lymphocyte reaction (MLR) declines with age in both mice (134) and men (135) as does also antibody-dependent T cell cytotoxicity (136) and cell-mediated cytotoxicity (CTL) (137).

The effects of aging on cell-mediated immune mechanisms are shown in Table 2.

Humoral Immunity

Infections caused by certain encapsulated bacteria, including *Streptococcus pneumoniae*, group B *Streptococcus*, and *Escherichia coli* K1, appear to occur more frequently in the elderly than in young adults (138, 139) and are suggeated to be due to a decline of humoral immune function that occurs during secescence. Although the total concentration of immunoglobulins remains constant, changes in serum immunoglobulin classes have been noted with age. Most reports reveal a gradual increase in the amount of IgA and IgG, whereas IgM concentrations are unchanged (91) and mortality was higher in a subgroup of volunteers with decreased IgG (140).

In contrast to age-associated alterations in T cell function, those involving B lymphocyte function or humoral immunity are relatively

Table 2. Effects of aging on cell-mediated immune mechanisms

Effect	Reference(s)
Thymic involution	1, 2, 4, 19–21, 24 ↓ 25
Thymic hormone levels T cell numbers	¥ 25
Total T cells	↓ 34–36
	→ 31–33
Immature T cells	↑ 37 ↑ 38 30
T helper cells	↑ 38, 39 ↓ 44, 45
F	\rightarrow 41, 42
	↑ 40, 41
T suppressor cells	$ \uparrow 40, 41 \downarrow 38, 42, 45, 63 \rightarrow 39, 43 $
	→ 39, 43 ↑ 32, 41
T cell responses	1 04, 11
Proliferative activity	↓ 39, 47–49, 52, 53
Suppressor activity	↓ 59–61, 63
	→ 56 ↑ 55, 56–58
Responses to IL-2	↓ 40, 65
•	→ 70
DTH	↓ 125
Resistance to infection MLR	↓ 126, 128, 131
Resistance to tolerance	↓ 126, 128, 131 ↓ 134, 135 ↓ 133
CTL in number and	↓ 131, 137
function	
Lymphokines	
IL-2 production	↓ 40, 65–69
IFN v production	→ 39 ↓ 72, 73, 128
IFN-γ production	$\begin{array}{c} 4 & 72, 73, 128 \\ \rightarrow & 71 \end{array}$
_	

 $[\]uparrow$, increased; \rightarrow , unchanged; \downarrow , decreased.

few. Investigations have demonstrated a decreased antibody response to the hepatitis B virus (141) and multivalent influenza vaccines (142, 143) in elderly individuals as well as an impaired adility to sustain the production of IgG when experimentally immunized with monomeric flagellin (144). When infected with influenza virus, elderly persons, particularly those with underlying diseases, are at increased risk for morbidity and mortality (145). In some studies the antibody response in the elderly to pneumococcal vaccine was sufficient to protect against infection (146, 147). Another study demonstrated the ability of the elderly to

Table 3. Effects of aging on humoral immune mechanisms

Effect	Reference(s)
Reduced antibody responses to:	
Specific antigens	149
Polyclonal activators	77
Vaccines	
Hepatitis B	141
Influenza	142, 143
Pneumococcal polysaccharide	146-148
Tetanus toxoid	151, 152
Enhanced autoantibody	154
Enhanced anti-idiotype antibody	149, 150
B cell function	
Primary defect	78, 79
Defect secondary to regulatory	
T cell abnormality	
Helper cells	113-116
Suppressor cells	63
Primary B cell defect and	39
regulatory T cell abnormality	

mount a polyclonal antibody response to pneumococcal polysaccharide vaccine that was similar to the response of healthy younger controls except for the IgM class responses, which were significantly weaker in the elderly (148).

It has been proposed that auto-anti-idiotypic antibodies that combine with surface immunoglobulin on B lymphocytes to inhibit antibody formation may be responsible for the alterations in the humoral immune response seen in senescence (149, 150).

The highest incidence of tetanus infection occurs in the elderly population. The mortality rate in persons over 65 years old approaches 80 per cent. The age-related decline found in both *in vivo* and *in vitro* synthesis of antitetanus toxoid antibody was suggested to be accounted for the impaired tetanus toxoid-specific T-helper cell activity as well as B-cell dysfunction (151, 152).

The effects of aging on humoral immune mechanisms are shown in Table 3.

Autoimmunity

It seems paradoxical that at the time in life

when the activity of the immune system is declining, the incidence of antoantibody production begins to rise. In aging mice an increased resistance to the induction of tolerance has been demonstrated (153). There is little direct evidence that the autoantibodies produced with advancing age have any deleterious effect. The role of antilymphocyte antibodies found with increasing frequency in the elderly is less certain (154). On the other hand, it has been postulated that low-grade tissue damage by a range of age-related autoantibodies may actually contribute to the process of senescence, although there is evidence to suggest that the production of autoantibodies in the elderly represents homeostatic control of the immune system.

Restoration of Immune Functions of the $$\operatorname{\textbf{Aged}}$$

Various attempts have been made to restore immune functions of aged animals to levels approaching those of younger mature individuals. Studies attempting to potentiate immune functions of old mice revealed that the loss of normal immune functions with age is associated with changes in antigen-/mitogenresponsive T cells, the inability of the involuted thymus to synthesize T cell maturation factor(s), changes in precursor cells in the bone marrow, and emergence of deleterious factors with age (6). When long-lived old mice were grafted with both young bone-marrow stem cells and newborn thymic lobes, their immune functions were restored to levels approaching those of younger adult mice, and the restorative effect was observed for 6 to 11 months after grafting in mice with a mean life span of 28 months (an equivalent of about 16-28 human years) (155).

The sulfhydryl compound most commonly used by immunologists is 2-mercaptoethanol (2-ME). Studies on its immunorestorative actions on aging mice show that it enhances the antibody-forming capacity of old mice pre-

ferentially over that of young mice (156). Thus, the effect of 2-ME on the T celldependent antibody-forming capacity of old spleen cells in vitro was an order of magnitude greater than that on young spleen cells (157). That 2-ME is also an effective immunorestorative agent in intact old mice was demonstrated by restoration of the T cell-dependent antibody-forming capacity of long-lived old mice to that of young mice by administration of this compound (156). More recently, in a preliminary study, young and old mice were subjected to immunotherapy by injecting either saline or dithiothreitol, a potent in vitro immunostimulant, following inoculation with melanoma cells (158). The results revealed that dithiothreitol could reduce the incidence of pulmonary metastasis 38 days after inoculation of melanoma cells. Moreover, augumentation of intracellular glutathione concentraions in lymphocytes may enhance immune function in depressed immune states (159). The mode of action of these chemicals is not known. It is well known that the function of sulfhydryl compounds ranges from R-SH to R-S-S-R' exchange reactions at the membrane level, to antioxidant and metal chelating effects (160).

Many recent reports point out the relationship between nutrition and immunocompetency in the elderly. The use of certain drugs such as cholestyramine, anticonvulsant drugs, and thiazide diuretics may reduce the immune system by inducing nutrient depletion (161). An important role is played by zinc depletion. Previous results have demonstrated that mice fed a zinc-supplemented diet maintain thymic hormone levels better than mice fed a normal diet (162). The effect mediated by zinc seems to be selective for B cells. In fact, zinc addition in culture augments specific antibody response or polyclonal antibody synthesis (163, 164). On the basis of the well-known capacity of zinc to activate B lymphocytes, the above-described effects are likely due to a modulation of early events involved in the activation of antibody forming cells.

The relation between zinc level and thymic hormones has also prompted many studies on the beneficial effects of thymic hormone administration in elderly individuals. In this context, it has been observed that injection of thymosin is able to restore T cell-dependent immune functions (165). With regard to the mechanism of action of thymosin, the enhancement of IL-2 production cannot be excluded (2).

Conclusion

In the present review, we have summarized the changes in immunocytes and discussed their functions in relation to the host defense network that are coupled with aging. It is proposed that increased susceptibility of elderly people to infectious and neoplastic diseases may be a consequence of immune senescence.

It is unlikely that all immune cells and systems age at equal rates. Although a plethora of frequently conflicting evidence has accumulated from studies in both animals and humans, the most visible cellular target of aging appears to be the T cells, and changes in their subpopulations involved in immunoregulation are highly prominent. These evidence appear to be related to thymic function which declines with age as assessed by a reduction in thymic hormone levels, thymic involution and its reduced activity. The functions of macrophages, PMN, NK cells, and also the complement system are not seriously impaired with age.

While it appears that impaired immune responsiveness is a consequence of the aging process, the possibility that altered immunity plays a primary role, if not wholly, in the senescence process remains to be solved. The evidence that lymphoid progenitors in bone marrow cells from young animals are able to differentiate into lymphoid cells in aging animals suggests that there may be a potential for reconstitution of some immune defects in aged

individuals.

A recent study suggested that in addition to a central role for immune mechanisms, the thymus appears to be closely related to the function of the endocrine system of the pituitary and the hypothalamus. In fact, involution of the thymus may not be irreversible but could be restored by manipulating the endocrine system (166). This hypothesis has led to the investigation of an effective manipulative methodology including grafting and treating with chemical agents by which immune system dysfunction might be prevented, retarded or restored in an effort to delay the inevitable consequences of age and age-related diseases.

ACKNOWLEDGMENT

We thank Mrs. Reiko Tanaka, nee Horikoshi, for assistance in the preparation of this manuscript.

REFERENCES

- 1) Ford PM. The immunology of ageing. Clin Rheum Dis 1986; **12**: 1–10.
- Antonaci S, Jirillo E, Bonomo L. Immunoregulation in aging. Diag Clin Immunol 1987; 5: 55–61.
- 3) Thoman ML, Weigle WO. The cellular and subcellular bases of immunosenescence. Adv Immunol 1989; **46**: 221–261.
- Saltzman, RL, Peterson PK. Immunodeficiency of the elderly. Rev Infect Dis 1987; 9: 1127–1139.
- 5) Busby J, Caranasos GJ. Immune function, autoimmunity, and selective immunoprophylaxis in the aged. Med Clin North Am 1985; **69**: 465–474.
- Makinodan T, Hirayama R. Age-related changes in immunologic and hormonal activities. IARC Sci Publ 1985; 58: 55–70.
- Peterson PK. Host defense abnormalities predisposing the patient to infection. Am J Med 1984; 76(Suppl 5A): 2–10.
- Dinarello CA. An update on human interleukin-1: From molecular bioloy to clinical relevance. J Clin Immunol 1985; 5: 287–297.
- 9) Gillis S. Interleukin 2: Biology and biochemistry. J Clin Immunol 1983; 3: 1-13.
- 10) Vilcek J, Gray PW, Rinderknecht E, Sevasto-

- poulos CG. Interferon-gamma: A lymphokine for all seasons. Lymphokines 1985; 11: 1–32.
- Kagaya K, Watanabe K, Fukazawa Y. Gapacity of recombinant gamma interferon to activate macrophages for Salmonella-killing activity. Infect Immun 1989; 57: 609–615.
- Nobholz M, MacDonald HR. Cytotoxic T lymphocytes. Ann Rev Immunol 1983; 1: 273–306.
- Mason DW, Morris PJ. Effector mechanisms in allograft refection. Ann Rev Immunol 1986; 4: 119–145.
- 14) Southern P, Oldstone MBA. Medical consequences of persistent viral infection. New Engl J Med 1986; 314: 359–367.
- Ortaldo JR, Herberman RB. Heterogeneity of natural killer cells. Ann Rev Immunol 1984;
 359–394.
- 16) Flesch I, Kaufmann SHE. Mycobacterial growth ingibition by interferon-γ-activated bone marrow macrophages and differential susceptibility among strains of Mycobacterium tuberculosis. J Immunol 1987; 138: 4408–4413.
- 17) Brummer E, Morrison CJ, Stevens DA. Recombinant and natural gamma interferon activation of macrophages in vitro: Different dose requirements for induction of killing activity against phagocytizable and nonphagocytizable fungi. Intect Immun 1985; 49: 724–730.
- 18) Peck R. Gamma interferon induces monocyte killing of *Listeria monocytogenes* by an oxygendependent pathway; alpha- or betainterferons by oxygen-independent pathways. J Leuk Biol 1989; 46: 434–440.
- Walford RL. Immunology and aging. Philip Levine Award. Am J Clin Pathol 1980; 74: 247–253.
- Steinmann GG, Klaus B, Muller-Hermelink H-K. The involution of the ageing human thymic epithelium is indepenent of puberty. Scand J Immunol 1985; 22: 563–575.
- Steinmann GG. Changes in the human thymus during aging. Cur Top Pathol 1986; 75: 43–88.
- Weksler ME, Siskind GW. The cellular basis of immune senescence. In: Suer HW ed. Monographs in developmental biology, vol 17. Basel: Karger, 1984: 110–121.
- Hirokawa K, Makinodan T. Thymic involution: Effect on T cell differentiation. J Immunol 1975; 114: 1659–1664.
- 24) Eren R, Zharhary D, Abel L, Globerson A.

- Age-related changes in the capcity of bone marrow cells to differentiate in thymic organ cultures. Cell Immunol 1988; **112**: 449–455.
- Lewis VM et al. Age, thymic involution and circulating thymic hormone activity. J Clin Endocrinol Metab 1978; 47: 145–150.
- 26) Ershler WB, Moore AL, Socinski MA. Influenza and aging: Age related changes and the effects of thymosin on the antibody response to influenza vaccine. J Clin Immunol 1984; 4: 445–454.
- 27) Zatz MM, Oliver J, Samuels C et al. Thymosin increases production of T-cell growth factor by normal human peripheral blood lymphocytes. Proc Natl Acad Sci USA 1984; 81: 2882–2887.
- 28) Pandolfi F, Quinti I, Montella F et al. T-dependent immunity in aged humans. II. Clinical and immunological evaluation after three months of administering a thymic extract. Thymus 1983; 5: 235–240.
- 29) Greenstein BD, Fitzpatrick FTA, Adcock IM et al. Reappearance of the thymus in old rats after orchidectomy: Inhibition of regeneration by testosterone. J Endocr 1986; 110: 417–422.
- 30) Fitzpatrick TA, Greenstein BD. Effects of various steroids on the thymus, spleen, ventral prostate and seminal vesicles in old orchidectomized rats. J Endocr 1987; 113: 51-55.
- 31) Charpentier B, Fournier C, Fries D *et al.* Immunological studies in human ageing 1. In vitro functions of T cells and polymorphs. J Clin Lab Immunol 1981; **5**: 87–93.
- 32) Kishimoto S, Tomino S, Inomata K, et al. Age-related changes in the subsets and functions of human T lymphocytes. J Immunol 1978; 121: 1773–1780.
- 33) Gupta S, Good RA. Sub-population of human T-lymphocytes. X. Alterations in T, B, third population cells and T cells with receptors for immunoglobulin M or G in aging humans. J Immunol 1979; 122: 1214–1219.
- 34) Clot J, Charmasson E, Brochier J. Agedependent changes of human blood lymphocyte subpopulations. Clin Exp Immunol 1972; 32: 346–351.
- 35) Diaz-Jouanen E, Strickland RG, Williams RC Jr. Studies of human lymphocytes in the newborn and the aged. Am J Med 1975; 58: 620–628.
- 36) Ales-Martinez JE, Alvarez-Mon M, Merino F et al. Decreased TcR-CD3⁺ T cell numbers in

- healthy aged humans. Evidence that T cell defects are masked by a reciprocal increase of TcR-CD3⁻CD2⁺ natural killer cells. Eur J Immunol 1988; **18**: 1827–1830.
- 37) Hallgren HM, Kersey JH, Dubey DP, Yunis EJ. Lymphocyte subsets and integrated immune function in aging humans. Clin Immunol Immunopathol 1978; 10: 65–78.
- 38) Moody CE, Innes JB, Staiano-Coico L et al. Lymphocyte transformation induced by autologous cells. XI. The effect of age on the autologous mixed lymphocyte reaction. Immunology 1981; 44: 431–438.
- 39) Rosenberg JS, Gilman SC, Feldman JD. Effects of aging on cell cooperation and lymphocyte responsiveness to cytokines. J Immunol 1983; 130: 1754–1758.
- Delafuente JC. Immunosenescence: clinical and pharmacologic consideration. Med Clin North Am 1985; 69: 475–486.
- 41) Chandra RK. Nutrition-immunity-infection interactions in old age. *In*: Chandra RK, ed. Nutrition, immunity and illness in the elderly. Oxford: Pergamon Press, 1985; 87–96.
- 42) Nagel JE, Chrest FJ, Adler WH. Enumeration of T lymphocyte subsets by monoclonal antibodies in young and aged humans. J Immunol 1981; 127: 2086–2088.
- 43) Quinti I, Pandolfi F, Fiorilli M et al. T-dependent immunity in aged humans. I. Evaluation of T-cell subpopulations before and after short term administration of a thymic extract. J Gerontol 1981; 36: 674–679.
- 44) Mascart-Lemone F, Delespesse G, Servais G, Kunstler M. Characterization of immunoregulatory T lymphocytes during aging by monoclonal antibodies. Clin Exp Immunol 1982; 48: 148–154.
- 45) Ligthart GJ, Corberand J, Fournier C et al. Admission criteria for immunogerontological studies in man: The Senieur Protocol. Mech Ageing Dev 1984; 28: 47–55.
- 46) Lightart GJ, Schuit HRE, Hijmans W. Subpopulations of mononuclear cells in ageing: Expansion of the null cell compartment and decrease in the number of T and B cells in human blood. Immunology 1985; 55: 15–21.
- 47) Hallgren HM, Buckley CE, Gilbertsen VA, Yunis EJ. Lymphocyte phytohemagglutinin responsiveness. Immunoglobulins and autoantibodies in aging humans. J Immunol 1973; 111: 1101–1107.
- 48) Murasko DM, Nelson BJ, Silver R et al. Immunologic response in an elderly popula-

- tion with a mean age of 85. Am J Med 1986; **81**: 612–618.
- 49) Hefton JM, Darlington GJ, Casazza BA, Weksler ME. Immunological studies of aging.
 5. Impaired poliferation of PHA responsive human lymphocytes in culture. J Immunol 1980; 125: 1007–1010.
- 50) Goodwin JS, Messner RP. Sensitivity of lymphocytes to prostaglandin E₂ increases in subjects over age 70. J Clin Invest 1979; 64: 434–439.
- Finkelstein MS. Unusual features of infections in the aging. Geriatrics 1982; 37: 65–78.
- 52) Nagel JE, Yanagihara RH, Adler WH. Cells of the immune response. *In*: Cristofalo VJ, Adelman RC, Roth GS, eds. CRC handbook of cell biology of aging. Boca Raton, Fla: CRC Press, 1985: 341–363.
- 53) Brock MA. Age-related changes in circannual rhythms of lymphocyte blastogenic responses in mice. Am J Physiol 1987: 252: R299–305.
- 54) Gershon R, Metzler C. Suppressor cells in aging. *In*: Makinodan T, Yunis E, eds. Immunology and Aging. New York: Plenum Press, 1977: 103–110.
- 55) Miller RD, Calkins CE. Development of selftolerance in normal mice. Appearance of suppressor cells that maintain adult selftolerance follows the neonatal autoantibody response. J Immunol 1988; 141: 2206–2210.
- 56) Callard RE, de St. Groth BF, Basten A, McKenzie, FC. The immune function in aged mice. V. Role of suppressor cells. J Immunol 1980; 124: 52–58.
- 57) Globerson A, Abel L, Barsilay M, Zan-Bar I. Immunoreglatory cells in aging mice. I. Concanavalin A-induced and naturally occurring suppressor cells. Mech Ageing Dev 1982; 19: 293–306.
- 58) Coria G, Mancini M, Adorini L. Immunoregulation in senescence: Increased inducibility of antigen specific suppressor T cells and loss of cell sensitivity to immunosuppression in aging mice. Proc Natl Acad Sci USA 1982; 79: 3803–3807.
- 59) Amagai T, Nakano K, Cinader B. Mechanisms involved in age-dependent decline of immune responsiveness and apparent resistance against tolerance induction in C57BL/6 mice. Scand J Immunol 1982; 16: 217–231.
- 60) Thoman ML, Weigle WO. Deficiency in suppressor T cell activity in aged animals. Reconstitution of this activity by interleukin 2. J Exp Med 1983; 157: 2184–2189.

- 61) Gottesman SRS, Walford RL, Thorbecke GJ. proliferative and cytotoxic immune functions in aging mice. II. Decreased generation of specific suppressor cells in alloreative cultures. J Immunol 1984; 133: 1782–1791.
- Doria G, Mancini C, Frasca D, Adorini L. Age restriction in antigen-specific immunosuppression. J Immunol 1987; 139: 1419–1425.
- 63) Kawanishi H, Kiely J. Immunoregulatory defects in murine aged Peyer's patches. Eur J Immunol 1987; 17: 1223–1228.
- 64) Gottesman SRS, Edington JM, Thorbecke GJ. Proliferative and cytotoxic immune functions in aging mice. IV. Effects of suppressor cell populations from aged and young mice. J Immunol 1988; 140: 1783–1790.
- 65) Gillis S, Kozak R, Durante M, Weksler ME. Immunological studies of aging: Decreased production of and response to T cell growth factor by lymphocytes from aged humans. J Clin Invest 1981; 67: 937–942.
- 66) Canonica GW, Caria M, Venuti D et al. T cell activation through different membrane structures (T3/Ti, T11, T44) and frequency analysis of proliferating and interleukin-2 producer T lymphocyte precursors in aged individuals. Mech Ageing Dev 1988; 42: 27–35.
- 67) Gilman SC, Rosenberg JS, Feldman JD. T lymphocytes of young and aged rats. II. Functional defects and the role of interleukine 2. J Immunol 1982; 128: 644–650.
- 68) Thoman ML, Weigle WO. Cell-mediated immunity in aged mice: An underlying lesion in IL-2 synthesis. J Immunol 1982; 128: 2358–2361.
- 69) Cheung HT, Twu J-S, Richardson A. Mechanism of the age-related decline in lymphocyte proliferation: Role of IL-2 production and protein synthesis. Exp Gerontol 1983; 18: 451–460.
- 70) Kennes B, Brohee D, Neve P. Lymphocyte activation in human ageing. V. Acquisition of response to T cell growth factor and production of growth factors by mitogen-stimulated lymphocytes. Mech Ageing Dev 1983; 23: 103–113.
- Miller AE. Selective decline in cellular immune response to varicella-zoster in the elderly. Neurology 1980; 30: 582–587.
- 72) Rytel MW, Larratt KS, Turner PA, Kalbfleisch JH. Interferon response to mitogens and viral antigens in elderly and young adult subjects. J Infect Dis 1986; 153: 984–987.
- 73) Abb J, Abb H, Deinhardt F. Age-related

- decline of human interferon alpha and interferon gamma production. Blut 1984; **48**: 285–289.
- 74) Buckler AJ, Vie H, Sonenshein GE, Miller RA. Defective T lymphocytes in old mice: Diminished production of mature c-myc RNA after mitogen exposure not attributable to alterations in transcription or RNA stability. J Immunol 1988; 140: 2442–2446.
- 75) Birkeland SA. Age-dependence of subpopulations and functions of human peripheral lymphocytes. J Clin Lab Immunol 1981; 5: 47–51.
- 76) Zharhary D. Age-related changes in the capability of the bone marrow to generate B cells. J Immunol 1988; 141: 1863–1869.
- 77) Pahwa SG, Pahwa RN, Good RA. Decreased in vitro humoral immune responses in aged humans. J Clin Invest 1981; 67: 1094–1102.
- 78) Hollingsworth JW, Gailotte R. B lymphocyte maturation in cultures from blood of elderly men: A comparison of plaque-forming cells, cells containing intracytoplasmic immunoglobulin and cell proliferation. Mech Ageing Dev 1981; 15: 9–18.
- 79) Whisler RL, Newhouse YG. Immunosenescence of the human B cell system: Impaired activation/proliferation in response to autologous monocytes pulsed with Staph protein A and the effects of interleukins 1 and 2 compared to interferon. Lymphokine Res 1985; 4: 331–337.
- 80) Weksler ME. Immune senescence in man. In: Fabris D, ed. Immunology and Ageing. The Hague: Martinus Nijhoff Publ., 1982: 165– 186.
- 81) Jones PG, Kauffman CA, Bergman AG *et al.* Fever in the elderly: production of leukocytic pyrogen by monocytes from elderly persons. Gerontology 1984; **30**: 182–187.
- 82) Gardner ID, Lim STK, Lawton JWM. Monocyte function in ageing humans. Mech Ageing Dev 1981; 16: 233–239.
- 83) Antonaci S, Jirillo E, Ventura MT et al. Non-specific immunity in ageing: Deficiency of monocyte and polymorphonuclear cellmediated functions. Mech Ageing Dev 1984; 24: 367–375.
- 84) Gardner ID, Remington JS. Aging and the immune response. II. Lymphocyte responsiveness and macrophage activation in *Toxoplasma gondii*-infected mice. J Immunol 1978; **120**: 944–949.
- 85) Mege JL, Capo C, Michel B et al. Phagocytic

- cell function in aged subjects. Neurobiol Ageing 1988; **9**: 217–220.
- 86) Weksler ME, Innes JB, Goldstein G. Immunological studies of aging. IV. The contribution of thymic involution to the immune deficiencies of aging mice and reversal with thymopoietin 32–36. J Exp Med 1978: 148: 996–1006.
- 87) Esposito, AL, Clark CA, Poirier WJ. An assessment of the respiratory burst and bactericidal activity of alveolar macrophages from adult and senescent mice. J Leuk Biol 1988; 43: 445–454.
- 88) Chang M-P, Makinodan T, Peterson WJ, Strehler BL. Role of T cells and adherent cells in age-related decline in murine interleukin 2 production. J Immunol 1982; 129: 2426–2430.
- Inamizu T, Chang M-P, Makinodan T. Decline in interleukin (IL)-1 production with age. Gerontologist 1983; 23: 249.
- Bradley SF, Vibhagool A, Kunkel SL, Kauffman CA. Monokine secretion in aging and protein malnutrition. J Leuk Biol 1989; 45: 510–514.
- 91) Phair JP. Host defense in the aged. *In*: Gleckman RA, Gantz NM, eds. Infections in the elderly. Boston: Little Brown, 1983: 1–12.
- Laharrague P, Corberand J, Fillola G et al. Impairment of polymorphonuclear functions in hospitalized geriatric patients. Gerontology 1983; 29: 325–331.
- Corberand J, Nguyen F, Laharrague P et al.
 Polymorphonuclear functions and aging in humans. J Am Geriatr Soc 1981; 29: 391–397.
- 94) Phair JP, Kauffman CA, Bjornson A *et al.* Host defenses in the aged: Evaluation of components of the inflammatory and immune responses. J Infect Dis 1978; **138**: 67–73.
- 95) Matsuyama SS, Jarvik LF, Fu TK, Kessler JO. Leukotaxis and serum immunoglobulins in the aged. Age 1981; **4**: 89–91.
- 96) Nagel JE, Han K, Coon PJ et al. Age differences in phagocytosis by polymorphonuclear leukocytes measured by flow cytometry. J Leuk Biol 1986; 39: 399–407.
- 97) Emmanuelli G, Lanzio M, Anfossi T et al. Influence of age on polymorphonuclear leukocytes in vitro: Phagocytic activity in healthy human subjects. Gerontology 1986; 32: 308–316.
- 98) Nagel JE, Pyle RS, Chrest FJ, Adler WH. Oxidative metabolism and bactericidal capacity of polymorphonuclear leukocytes from

- normal young and aged adults. J Gerontol 1982; **37**: 529–534.
- 99) Van Epps DE, Goodwin JS, Murphy S. Agedependent variations in polymorphonuclear leukocyte chemiluminescence. Infect Immun 1978; 22: 57–61.
- 100) Lipschitz DA, Udupa KB, McClellan JL. Protein kinase C (PKC) and the decreased response of neutrophils from the aged. Clin Res 1986; 34: 463A.
- 101) Fox RA. The effect of ageing on the immune response. *In*: Fox RA, ed. Immunology and infection in the elderly. New York: Churchill Livingstone, 1984: 289–309.
- 102) Itoh H, Abo T, Sugawara S et al. Age-related variation in the proportion and activity of murine liver natural killer cells and their cytotoxicity against regenerating hepatocytes. J Immunol 1988; 141: 315–323.
- 103) Fernandes G, Gupta S. Natural killing and antibody-dependent cytotoxicity by lymphocyte subpopulations in young and aging humans. J Clin Immunol 1981; 1: 141–148.
- 104) Nagel JE, Collins GD, Adler WH. Spontaneous or natural killer cytotoxicity of K562 erythroleukemic cells in normal patients. Cancer Res 1981; 41: 2284–2288.
- 105) Murasco DM, Nelson BJ, Silver R et al. Immunologic response in an elderly population with a mean age of 85. Am J Med 1986; 81: 612–618.
- 106) Batory G, Benczur M, Garam VT *et al.* Increased killer cell activity in aged humans. Immunobiology 1981; **158**: 393–402.
- 107) Onsrud M. Age dependent chambers in some human lymphocyte sub-populations. Acta Pathol Microbiol Scand Sec C 1981; 89: 55–62.
- 108) Krishnaraj R, Blandford G. Age-associated alterations in human natural killer cells. 1. Increased activity as per conventional and kinetic analysis. Clin Immunol Immunopath 1987; 45: 268–285.
- 109) Krishnaraj R, Blandford G. Age-associated alterations in human natural killer cells. 2. Increased frequency of selective NK subsets. Cell Immunol 1988; 114: 137–148.
- 110) Ligthart GJ, van Vlokhoven PC, Schuit HRE, Hijmans W. The expanded null cell compartment in ageing: increase in the number of natural killer cells and changes in T-cell and NK-cell subsets in human blood. Immunology 1986; 59: 353–357.
- 111) MacLean LD, Meakins JL, Taguchi K et al.

- Host resistance in sepsis and trauma. Ann Surg 1975; **182**: 207–217.
- 112) Yoshikawa TT. Aging and infectious diseases: state of the art. Gerontology 1984; 30: 275–278.
- 113) Liu JL, Segre M, segre D. Changes in suppressor, helper, and B-cell functions in aging mice. Cell Immunol 1982; 66: 372–382.
- 114) Staniano-Coico L, Darzynkiewicz Z, Melamed MR, Weksler ME. Immunological studies of aging. IX. Impaired proliferation of T lymphocytes detected in elderly humans by flow cytometry. J Immunol 1984; 132: 1788–1792.
- 115) Morgan EL, Weigle WO. The immune response in aged C57BL/6 mice. II. Characterization and reversal of a defect in the ability of aged spleen cells to respond to the adjuvant properties of Fc fragments. J Immunol 1982; 129: 36–45.
- Vissinga CS, Dirven CJAM, Steinmeyer FA et al. Deterioration of cellular immunity during aging: The relationship between age-dependent impairment of delayed-type hypersensitivity reactivity, interleukin-2 production capacity, and frequency of Thy-1+, Lyt-2⁻ cells in C57BL/Ka and CBA/Rij mice. Cell Immunol 1987; 108: 323–334.
- 117) Ledbetter JA, Herzenberg LA. Xenogeneic monoclonal antibodies to mouse lymphoid differentiation antigens. Immunol Rev 1979; 47: 63–90.
- 118) Gorczynski RM, Kennedy M, Macrae S. Alteration in lymphocyte recognition repertoire during aging. II. Changes in the expressed T-cell receptor repertoire in aged mice and the persistence of that change after transplantation to a new differentative environment. Cell Immunol 1983; 75: 226–241.
- 119) Blankwater MJ. Thesis, State University Utrecht. The Netherlands, 1978.
- 120) Segre M, Segre, D. Humoral immunity in aged mice. I. Age-related decline in the secondary response to DNP of spleen cells propagated in diffusion chambers. J Immunol 1976; 166: 731–738.
- 121) Ernst DN, Lubaroff DM. Membrane antigen phenotype of sensitized T lymphocytes mediating tuberculin delayed hypersensitivity in rats. Cell Immunol 1984; 88: 436–452.
- 122) Fukazawa Y, Kagaya K, Ishibashi Y. Effect of delayed-type hypersensitivity reaction and transferred lymphokine on the resistance of mice to *Salmonella typhimurium* infection. Infect Immun 1983; **39**: 986–989.

- 123) Mielke MA, Ehlers S, Hahn H. T-cell subsets in delayed-type hypersensitivity, protection, and granuloma formation in primary and secondary *Listeria* infection in mice: Superior role of Lyt-2⁺ cells in acquired immunity. Infect Immun 1988; **56**: 1920–1925.
- 124) Kagaya, K, Shinoda T, Fukazawa Y. Murine defense mechanism against *Candida albicans* infection. I. Collaboration of cell-mediated and humoral immunities in protection against systemic *C. albicans* infection. Microbiol Immunol 1981; 25: 647–654.
- 125) Marrie TJ, Johnson S, Durant H. Cell-mediated immunity of healthy adult Nova Scotians in various age group compared with nursing home and hospitalized senior citizens. J Allergy Clin Immunol 1988; 81: 836–844.
- 126) Orme IM. Aging and immunity to tuberculosis: Increased susceptibility of old mice reflects a decreased capacity to generate mediator T lymphocytes. J Immunol 1987; 138: 4414–4418.
- 127) Zurcher C, van Zwieten MJ, Solleveld HA, Hollander CF. *In*: Foster HK, Smal JD, Fox JG, eds. The mouse in biomedical research. Orlando: Academic Press, 1982: 11–35.
- 128) Rytel MW. Effect of age on viral infections: Possible role of interferon. J Am Geriatr Soc 1987; **35**: 1092–1099.
- 129) Hayward AR, Herberger M. Lymphocyte responses to varicella zoster virus in the elderly. J Clin Immunol 1987; 7: 174–178.
- 130) Hayward AR, Pontesilli O, Herberger M et al. Specific lysis of varicella zoster virus infected B lymphoblasts by human T cells. J Virol 1986; 58: 179–184.
- 131) Hayward AR, Herberger M, Laszlo M. Cellular interactions in the lysis of varicella zoster infected human fibroblasts. Clin Exp Immunol 1986; 63: 141–146.
- 132) Berger R, Florent G, Just M. Decrease of the lymphoproliferative response to varicella zoster virus antigen in the aged. Infect Immun 1981; **32**: 24–27.
- 133) Mayumi H, Good RA. Dependency of cyclophosphamide-induced skin allograft tolerance on age of adult recipient mice. Transplantation 1988; **46**: 451–453.
- 134) Gershon H, Merhav S, Abraham C. T-cell division and aging. Mech Ageing Dev 1979; 9: 27–38.
- 135) Goodwin JS, Searles RP, Tung KSK. Immunological response of a healthy elderly population. Clin Exp Immunol 1982; 48:

- 403-410.
- 136) Haffer K, Freeman MJ, Watson RR. Effects of age on cellular immune responses in Balb/CJ mice. Mech Ageing Dev 1979; 11: 279–285.
- 137) Miller RA. Age-associated decline in precursor frequency for different T cell-mediated reactions, with preservation of helper or cytotoxic effect per precursor cell. J Immunol 1984; 132: 63–68.
- 138) Verghese A, Berk SL. Bacterial pneumonia in the elderly. Medicine 1983; **62**: 271–285.
- 139) Gallagher PG, Watanakunakorn C. Group B streptococcal bacteremia in a community teaching hospital. Am J Med 1985; **78**: 795–800.
- 140) Buckley CE III, Buckley EG, Dorsey FC. Longitudinal changes in serum immunoglobulin levels in older humans. Fed Proc 1974; 33: 2036–2039.
- 141) Denis F, Mounier M, Hessel L et al. Hepatitis-B vaccination in the elderly. J Infect Dis 1984; 149: 1019.
- 142) Centers for Disease Control. Outbreaks of influenza among nursing home residents-Connecticut, United States. MMWR 1985; 34: 478–482.
- 143) Levine M, Beattie BL, McLean DM et al. Characterization of the immune response to trivalent influenza vaccine in elderly men. J Am Geriatr Soc 1987; 35: 609-615.
- 144) Berger R, Florent G, Just M. Decrease of the lymphoproliferative response to varicellazoster virus antigen in the aged. Infect Immun 1981; 32: 24–27.
- 145) Schneider EL. Infectious diseases in the elderly. Ann Intern Med 1983; **98**: 395–400.
- 146) Shapiro ED, Clemens JD. A controlled evaluation of the protective efficacy of pneumococcal vaccine for patients at high risk of serious pneumococcal infections. Ann Intern Med 1984; 101: 325–330.
- 147) Ammann AJ, Schiffman G, Austrian R. The antibody responses to pneumococcal capsular polysaccharides in aged individuals. Proc Soc Exp Biol Med 1980; 164: 312–316.
- 148) Ruben FL, Uhrin M. Specific immunoglobulin-class antibody responses in the elderly before and after 14-valent pneumococcal vaccine. J Infect Dis 1985; 151: 845–849.
- 149) Szewczuk MR, Campbell RJ. Loss of immune competence with age may be due to auto-antiidiotypic antibody regulation. Nature 1980; 286: 164–166.
- 150) Tsuda T, Kim YT, Siskind DW, Weksler ME.

- Old mice recover the ability to produce IgG and high-avidity antibody following irradiation with partial bone marrow shielding. Proc Natl Acad Sci USA 1988; 85: 1169–1173.
- 151) Kishimoto S, Tomino S, Mitsuya H *et al.* Age-related decline in the in vitro and in vivo synthesis of antitetanus toxoid antibody in humans. J Immunol 1980; **125**: 2347–2352.
- 152) Kraft R, Bachmann M, Bachmann K et al. Satisfactory primary tetanus antitoxin responses but markedly reduced germinal centre formation in first draining lymph nodes of ageing mice. Cell Exp Immunol 1987; 67: 447–453.
- 153) Goidl EA, Innes JB, Weksler ME. Immunological studies of aging. II. Loss of IgG and high avidity plaque forming cells and increased suppressor cell activity in aging mice. J Exp Med 1976; 144: 1037–1048.
- 154) Lokhorst HM, Vanderlinden JA, Schuurman HJ et al. Immun function during aging in man: Relation between serological abnormalities and cellular immune status. Eur J Clin Invest 1983; 13: 209–214.
- 155) Hirokawa K, Sato K, Makinodan T. Restoration of impaired immune functions in aging animals. V. Long-term immunopotentiating effects of combined young bone marrow and newborn thymus grafts. Clin Immunol Immunopathol 1982; 22: 297–304.
- 156) Makinodan T, Albright JW. Restoration of impaired immune functions in aging animals. III. Effect of mercaptoethanol in enhancing the reduced primary antibody responsiveness in vivo. Mech Ageing Dev 1979; 11: 1–8.
- 157) Makinodan T, Albright JW. Restoration of impaired immune functions in aging animals.
 II. Effect of mercaptoethanol in enhancing the reduced primary antibody responsiveness in vitro. Mech Ageing Dev 1979; 10: 325–340.
- 158) Hirayama R, Sato K, Makinodan T. The potentiating effect of dithiothreitol (DTT), an immunostimulant, on the resistance to pulmonary metastasis of melanoma cells in young and old mice. *In*: Proceedings of the 5th International Congress of Immunology, Kyoto, Japan, 1983.
- 159) Fidelus RK, Tsan MF. Glutathione and lymphocyte activation: a function of ageing and auto-immune disease. Immunology 1987; **61**: 503–508.
- 160) Broome JD, Jeng MW. Promotion of replication in lymphoid cells by specific thiols and disulfides in vitro. J Exp Med 1973; 138:

- 574-592.
- 161) Rikans LE. Drugs and nutrition in old age. Life Sci 1986; 39: 1027–1036.
- 162) Iwata T, Incefy GS, Tanaka T et al. Circulating thymic hormone levels in zinc deficiency. Cell Immunol 1979; 47: 100–105.
- 163) Malave I, Claverie-Benureau S, Benaim IR. Modulation by zinc of the in vitro antibody response to T-dependent and T-independent antigens. Immunol Commun 1983: 12: 397–406.
- 164) Winchurch RA, Thomas DJ, Adler WH,

- Lindsay TJ. Supplemental zinc restores antibody formation in cultures of aged spleen cells. J Immunol 1984; **133**: 569–571.
- 165) Goldstein AL, Low TLK, Thurman GB et al. Current status of thymosin and other hormones of the thymus gland. In: Greep RO, ed. recent progress in hormone research. New York: Academic Press, 1981: 369–415.
- 166) Utsuyama M, Hirokawa K. Hypertrophy of the thymus and restoration of immune functions in mice and rats by gonadectomy. Mech Ageing Dev 1989; 47: 175–185.